

		20	·/x		
1317		Genotypi	ing Report	narm.	
Strain ID	T007851	Strain Type	CKO(Cas9)	Genetic Background	C57BL/6JGpt
Designer	Ya'nan Xu	Gene Name	34×	Map3k7	, C
. Strategy of (Genotyping		Ъ.	arn _{ate}	
_//	F1 → ①5'arm	R1 ◀ ► Flox		'arm €	
	Loxp 🔶 Prim	er (1) (2) PC	R Reaction Number	- Sequencing Region	

Wild type: ①PCR reaction obtains a single WT band; ②PCR reaction obtains a single WT band. Heterozygote: ①PCR reaction obtains a WT band and a Targeted band; ②PCR reaction obtains a WT band and a Targeted band. Homozygote: ①PCR reaction obtains a single Targeted band; ②PCR reaction obtains a single Targeted band.

Note: The sizes of WT and Targeted band are shown below.

2. Primer Information

PCR No.	Primer No.	Sequence	Band Size	
(1)(5'arm)	T007851-F1	CTTTGGGAAGTGATCGACCCAG	WT:359bp	
	T007851-R1	GCATTGAGAGAGTGCTCTGCCC	Targeted:464bp	
@(3'arm)	T007851-F2	CAACTTATCCAGAGCGGTGCG	WT:368bp Targeted:469bp	
	T007851-R2	T007851-R2 GCAGATCAGAGCAAAGGCACAG		

3. Gel Image & Conclusion





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Note: P: Positive control; WT: Wildtype control; B: Blank control (ddH₂O); M: DNA Ladder

① Control (WT) : It is an important reference mark for whether the PCR reaction is successful and whether the product band position and size meet the theoretical requirements.

(2) Control (B) : PCR amplification was performed without template in the PCR reagent to monitor whether the reagent was contaminated.

4. PCR Condition

PCR Reaction Co	omponent	AX Con	20 ²	
Seg.	reactio	reaction component		
1 3/2	2 × Rapid Taq Master Mix (Va	2 × Rapid Taq Master Mix (Vazyme P222)		
2	ddH2O	The star	9.5	
3	Primer A(10pmol/µl)	Primer A(10pmol/µl)		
4 🖓	Primer B(10pmol/µl)	Primer B(10pmol/µl)		
5	Template(≈100ng/µl)	Template(≈100ng/µl)		
PCR program (1)	priority selection	m the		
Seg.	Temp.	Time	Cycle	
1 6	95°C	5min		
2	98°C	30s	20×	
3	65℃*(-0.5℃/cycle)	30s	2/2 3/2	
4	72℃	45s*	12× 4	
5 %	98°C	30s	20×	
6	55℃*	30s		
7 7	72℃	45s*	Ph Str	
8	72℃	5min	134 24	
9 7	10°C	hold	in the second se	
PCR program ②	the second choice			
Seg.	Temp.	Time	Cycle	
1	95°C	5min	197 ₀ 06	
2	98°C	30s	35×	
3	58°C*	30s 🔾		
4	72°C	45s*		
5	72°C	5min	19/2	
6 6	10°C	hold	97	

Note*: Annealing temperature and extension time can be determined according to the actual amplification situation and amplification enzyme efficiency.



