

		Genotyp	ing Report		
Strain ID	T020360	Strain Type	CKO(Cas9)	Genetic Background	C57BL/6JGpt
Designer	Ya'nan Xu	Gene Name	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	Slc2a10	Co Co
Strategy of C	Genotyping	73	1732	STEWS	
	F1 → ①5'arm	R1 Flox		'arm <b><sup>R2</sup></b>	
	Loxp 🔶 Prim	er (1) (2) PC	R Reaction Number	- Sequencing Region	

Wild type: ①PCR reaction obtains a single WT band; ②PCR reaction obtains a single WT band. Heterozygote: ①PCR reaction obtains a WT band and a Targeted band; ②PCR reaction obtains a WT band and a Targeted band. Homozygote: ①PCR reaction obtains a single Targeted band; ②PCR reaction obtains a single Targeted band.

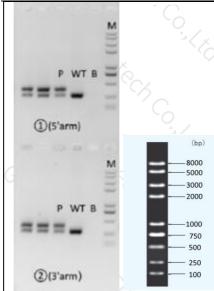
Note: The sizes of WT and Targeted band are shown below.

## 2. Primer Information

PCR No.	Primer No.	Sequence	Band Size
(1)(5'arm)	T020360-F1	GGATCTCTAAGTTCAAGGCCAGTC	WT: 271bp
	T020360-R1	TCAGGGTTACTTGCCAGCATTC	Targeted: 376bp
@(3'arm)	T020360-F2	GCTGGTCGTTACCGAGATAGGATC	WT: 289bp
	T020360-R2 GGCAAGGTCACATAGCATCCCT		Targeted: 395bp

## 3. Gel Image & Conclusion





Note: P: Positive control; WT: Wildype control; B: Blank control (ddH<sub>2</sub>O); M: DNA Ladder
① Control (WT) : It is an important reference mark for whether the PCR reaction is successful and whether the product band position and size meet the theoretical requirements.
② Control (B) : PCR amplification was performed without template in the PCR reagent to monitor whether the reagent was contaminated.

## 4. PCR Condition

PCR Reaction	Component	×	34 34	
Seg.	reaction co	Volume (µl)		
1 0	2 × Rapid Taq Master Mix(Vazym	2 × Rapid Taq Master Mix (Vazyme P222)		
2 7	ddH2O	str.	9.5	
3	Primer A(10pmol/µl)	Primer A(10pmol/µl)		
4	Primer B(10pmol/μl)	Primer B(10pmol/µl)		
5 %	Template(≈100ng/µl)	ate ch	1	
PCR program	① priority selection	No No	5. 6	
Seg.	Temp.	Time	Cycle	
1	95°C	5min	132001	
2	98°C	30s	20×	
3 %	65℃*(-0.5℃/cycle)	30s		
4	72℃ 45s*			
5	98°C 30s		20×	
6	55℃*	30s	nate a	
6		30s 45s*	nate ch	



9	10°C	a la	hold	100	( <sup>(</sup> C
PCR program	② the second choice	n n n n n n n n n n n n n n n n n n n	$\langle \varphi \rangle$		dra sla
Seg.	Temp.		Time		Cycle
1 200	<b>95°</b> C	6	5min	Ceno.	°°%
2	98°C	no.	30s	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	35×
3	58°C*	120	30s		m it
4	<b>72</b> ℃	3	45s*	3	
5 2	<b>72</b> ℃	G <sub>C</sub>	5min	ns,	20
6 dr	10°C		hold	2	
	2	- Phi	3/5		$\mathcal{T}_{\mathcal{A}}$

Note\*: Annealing temperature and extension time can be determined according to the actual amplification situation and amplification enzyme efficiency.