

- Phal	n Colle	Genoty	oing Report	nohanna.	College College
Strain ID	T055220	Strain Type	KI(Cas9)	Genetic Background	C57BL/6JGpt
Designer	Tianjiao Wang	Gene Name	HII	-CAG-LSL-Hoxa5-HA-Po	lyA
1. Strategy of	f Genotyping	- A	n.	C <sup>arma</sup> te	
_//_		<sup>F1</sup> 1)5'arm	R1		_//

Primer 12

F2

Sequencing Region PCR Reaction Number

R2

5-C0-44

Wild type: ①PCR reaction obtains none band; ②PCR reaction obtains a WT band. Heterozygote: ①PCR reaction obtains a Targeted band; ②PCR reaction obtains a WT band. Homozygote: ①PCR reaction obtains a Targeted band; ②PCR reaction obtains none band. Note: The sizes of WT and Targeted band are shown below. For 2PCR reaction, because the WT band is much smaller than the target band, it is likely to produce dominant amplification, the reaction is only used to judge whether there is a WT allele.

(2)WT

## 2. Primer Information

PCR No.	Primer No.	Sequence	Band Size	
(1)5'arm	T055220-F1	GGGCAGTCTGGTACTTCCAAGCT	WT:0bp	
	T055220-R1	TCAATGGAAAGTCCCTATTGGCGT	Targeted:353bp	K.
OWT	T055220-F2	AGTCTTTCCCTTGCCTCTGCT	WT:825bp Targeted:5325bp	8
2WT	T055220-R2	GGGTCTTCCACCTTTCTTCAG		0

## 3. Gel Image & Conclusion

Colley





Note: P:Positive control; WT: Wildype control; B: Blank control (ddH<sub>2</sub>O); M: DNA Ladder
① Control (WT) : It is an important reference mark for whether the PCR reaction is successful and whether the product band position and size meet the theoretical requirements.
② Control (B) : PCR amplification was performed without template in the PCR reagent to monitor whether the reagent was contaminated.

## 4. PCR Condition

2 d 3 P 4 P 5 T PCR program ① priori Seg. T 1 9	· · · · · · · · · · · · · · · · · · ·	7	Volume (μl) 12.5 9.5 1 1 1
2 d 3 P 4 P 5 T PCR program ① priori Seg. T 1 9	dH2O rimer A(10pmol/μl) rimer B(10pmol/μl) emplate(≈100ng/μl) <b>ty selection</b>		9.5
3         P           4         P           5         Ti           PCR program ① priori         Seg.           1         9	rimer A(10pmol/µl) rimer B(10pmol/µl) emplate(≈100ng/µl) <b>ty selection</b>		
4 Pr 5 Tr PCR program ① priori Seg. Tr 1 9	rimer B(10pmol/µl) emplate(≈100ng/µl) ty selection		1
5 Ti PCR program ① priori Seg. Ti 1 9	emplate(≈100ng/μl) ty selection		- "OX"
PCR program ① priori Seg. Tu 1 9	ty selection	C C	1
Seg. Ti	· · · · · · · · · · · · · · · · · · ·	- 10	
1. 2. 2.		~./. 12	~~~~~
<u> </u>	emp.	Time	Cycle
	5°C 975	5min	100 CC
2 9	8°C	30s	20× 5
3 6	5℃*(-0.5℃/cycle)	30s	G
4 7. 7.	2°C 🗘 🂫	45s*	B. Ka
5 9	8°C	30s	20×
6 5	5°C* 6	30s	10
7 2 7	2°°4 / 20,	45s*	
8 7	2°C	5min	20, 20
9	0°C 20	hold	130
PCR program ② the s	econd choice		7. A.K.
Seg. To	emp.	Time	Cycle
1 9	1/5	5min	A. (A)



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2	110h.	98°C 😪	100	30s	20	35×
3	910	58°C*	13y	30s 🔗		an sh
4	0 <sup>(2</sup> )	72℃	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	45s*	~	
5	Ch.	72℃	Ċ.	5min	Cox.	19 A
6	200	10°C 🤇	CO.	hold	- 10	6

Note\*: Annealing temperature and extension time can be determined according to the actual amplification situation and amplification enzyme efficiency.

